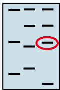
QuickProtocol™

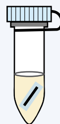
ISO 9001 ISO 14001

GeneJET™ Gel Extraction Kit, #K0691, #K0692

Note. All steps should be carried out at room temperature. All centrifugations should be carried out in a microcentrifuge at ≥12 000 x g (10 000-14 000 rpm, depending on the rotor type).

1





Prepare the DNA for binding

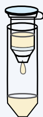
Excise the gel slice with the DNA fragment and weigh.

Add a 1:1 volume of **Binding Buffer** to the gel slice (volume/weight) (e.g. add 100 µl of Binding buffer for every 100 mg of agarose gel).

Incubate the gel mixture at 50-60°C for 10 min. Invert the tube every few min.

Note:
- For gels with an agarose content ≥ 2%, add 2:1 volumes of Binding Buffer to the gel slice.
- If the DNA fragment is ≤500 bp or >10 kb, *see* complete protocol in the manual for additional recommendations.

2



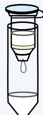
Bind DNA

Transfer the solution to the GeneJET™ Purification Column.

Centrifuge for 30-60 s. Discard the flow-through.

Note: If the purified DNA will be used for sequencing *see* complete protocol in the manual for additional recommendations.

3



Wash the column

Add 700 µl of **Wash Buffer** and centrifuge for 1 min.

Discard the flow-through.

Centrifuge empty column for 1 min.

4

Elute purified DNA

Place the column into a fresh 1.5 ml microfuge tube.

Add 50 µl of **Elution Buffer** to the column.

Centrifuge for 1 min. Collect the flow-through.

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