

NZY5 α Competent Cells

Catalogue number:

MB00401 (20 transformations)

MB00402 (40 transformations)

Description

NZY5 α Competent Cells display similar properties to DH5 α , which are suitable for high efficiency transformation in a wide variety of applications. The ϕ 80d/*lacZ* Δ M15 marker provides α -complementation of the β -galactosidase gene from pUC or similar vectors and, therefore, can be used for blue/white screening of colonies on bacterial plates containing Blue-gal or X-gal. NZY5 α can also serve as a host for the M13mp cloning vectors. In addition to supporting blue/white screening, *recA1* and *endA1* mutations in DH5 α TM Cells increase insert stability and improve the quality of plasmid DNA prepared from minipreps.

Genotype: *fhuA2* Δ (*argF-lacZ*)U169 *phoA glnV44* ϕ 80 Δ (*lacZ*)M15 *gyrA96 recA1 relA1 endA1 thi-1 hsdR17*

Storage temperature

The NZY5 α Chemically Competent *Escherichia coli* cells are shipped on dry ice. Upon receipt, store at -80 °C.

System Components

NZY5 α Competent Cells (10 or 20 \times 200 μ L)

Competent Cells Control Plasmid (10 μ L at 0.1 ng/ μ L)

Transformation Protocol

1. A Competent cells control plasmid solution (0.1 ng/ μ L) is provided as a control to determine the transformation efficiency. To obtain maximum transformation efficiency, the experimental DNA must be free of phenol, ethanol, protein and detergents.
2. Thaw competent cells on ice. Gently mix cells. Do not mix cells by pipetting.
3. To determine the transformation efficiency, add 1 μ L of a $\frac{1}{10}$ dilution of control plasmid DNA (0.01 ng) to one tube containing 100 μ L competent cells. Move the pipette through the cells while dispensing. Gently tap tube to mix.
4. For DNA from ligation reactions, add 5 to 10 μ L of the reaction (10 to 100 ng DNA) to the NZY5 α competent cells. Gently tap tubes to mix.

5. Incubate cells on ice for 30 minutes.
6. Heat-shock cells for 40 seconds in a 42 °C water bath; do not shake.
7. Place on ice for 2 minutes.
8. Add 0.9 mL room temperature SOC Medium.
9. Shake at 225 rpm (37 °C) for 1 hour.
10. Spread 50 to 150 μ L of cells transformed with competent cells control plasmid on LB agar plates containing 100 μ g/mL ampicillin.
11. Spread 100 to 250 μ L of the cells transformed with the ligation reaction on LB agar plates containing the required antibiotic and, if required, X-Gal. To obtain the maximum number of colonies, spin the 1000 μ L cell culture for 1 min at 5000 rpm, remove 800 μ L of media and spread cells after re-suspending in the remaining buffer.
12. Incubate overnight at 37 °C.

Notes

1. Competent cells are very sensitive to changes in temperature and should be thawed on ice. The transformation should be started immediately after the cells are thawed. For best results, each vial of cells should be thawed only once. Although the cells are re-freezable, subsequent freeze-thaw cycles will lower transformation frequencies by approximately two-fold.
2. Competent cells must be treated gently. Mix cells by swirling or gently tapping the reaction tube. Do not mix by pipetting or vortexing.
3. Media other than SOC can be used, but the transformation efficiency will be reduced. Using LB reduces transformation efficiency a minimum of two- to three-fold.
4. Transformation efficiencies will be approximately 10-fold lower for ligation of inserts to vectors than for an intact control plasmid.

Quality Control:

NZY5 α Competent Cells consistently yield $> 1.0 \times 10^9$ colony-forming units/ μ g competent cells control plasmid when transformed with non-saturating amounts of DNA (0.01 ng/100 μ L cells).

Certificate of Analysis

Test	Result
> 1.0×10^9 of colony-forming units/ μg competent cells control plasmid	Pass
Cells contamination	Pass

Approved by:



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