

# NZY T7 High Yield RNA Synthesis kit

## Catalogue number:

MB36301, 50 reactions

## Description

NZY T7 High Yield RNA Synthesis kit was designed for the *in vitro* production of large amounts of RNA transcripts from any DNA template subjected to the control of a T7 promoter sequence. This kit was formulated to provide high yields of RNA transcripts. Yields of approximately 150-200 µg of RNA from 1 µg DNA template may be expected from a 2 hours reaction at 37 °C. The resulting RNA transcripts present high integrity and the required quality for a variety of downstream applications, such as RNA splicing studies or RNA-protein interaction experiments.

The *in vitro* transcription reaction is performed by an engineered version of phage T7 RNA polymerase. This enzyme displays a high specificity for the T7 promoter allowing an effective transcription of the DNA strand of interest from the promoter sequence. NZY T7 High Yield RNA Synthesis kit can be used to transcribe linearized plasmid DNA templates or DNA templates generated by PCR.

To maintain integrity of generated RNA transcripts, the T7 HY Enzyme Mix includes a Ribonuclease Inhibitor protein to effectively protect synthesised RNA transcripts from degradation by contaminating RNases resulting from different sources, such as plasmid templates or others.

## Storage temperature

Store all kit components at -20 °C in a freezer without defrost cycles. Stability can be extended by storage at -80 °C. NZY T7 High Yield RNA Synthesis kit components are stable up to the expiry date specified with the product.

## Kit components

Component	Amount
T7 HY Enzyme Mix	100 µL
10× HY Transcription Reaction Buffer	100 µL
100 mM ATP	50 µL
100 mM CTP	50 µL
100 mM GTP	50 µL
100 mM UTP	50 µL
Linear DNA Control <sup>a</sup> (150 ng/µL)	10 µL

<sup>a</sup> Linear DNA Control: linear vector provided for 3 experiments.

## Protocol for high yield *in vitro* RNA synthesis

NZY T7 High Yield RNA Synthesis kit was optimized to produce large amounts of RNA transcripts using unmodified ribonucleotides. Nevertheless, this kit is compatible with capped RNA synthesis or RNA synthesis using modified ribonucleotides (please read the support protocols available at [www.nzytech.com](http://www.nzytech.com)). The standard protocol for *in vitro* transcription is presented below.

### Standard *in vitro* Transcription

Before you start using this protocol, please read carefully the Important Notes section.

After mixing and quickly pulsing the reaction components, place T7 HY Enzyme Mix and ribonucleotides solutions on ice but **keep the 10× HY Transcription Reaction Buffer at room temperature**.

*Note: Some components of 10× HY Transcription Reaction Buffer can co-precipitate the template DNA if the reaction is assembled on ice.*

1. Set up the transcription reaction at **room temperature** by adding the below mentioned components in the order presented in the following table.

Nuclease-free water	up to 20 µL
10× HY Transcription Reaction Buffer	2 µL
100 mM ATP	1 µL
100 mM CTP	1 µL
100 mM GTP	1 µL
100 mM UTP	1 µL
Template DNA*	0.5-1 µg
T7 HY Enzyme Mix	2 µL

\* For control reaction, use 0.5 µg of Linear DNA control (3 µL). The control template DNA is a 6043 bp linearized plasmid which codes for a 600 nt RNA transcript.

2. Mix thoroughly and pulse the reaction to collect all drops.
3. Incubate at 37 °C for 2 hours.  
*Note: For short transcripts (<0.3 Kb), incubate the *in vitro* transcription reactions at 37 °C for 4 hours or more.*

### 4. *Optional: DNase treatment to remove DNA template*

If the template DNA will interfere with the downstream application of the RNA transcript, proceed with a DNase digestion directly after the *in vitro* transcription reaction is terminated.

Add DNase (RNase-free) plus DNase reaction buffer to the transcription reaction, mix and incubate for 15 minutes at 37 °C. Stop DNase activity by adding 1 µL of 0.5 M EDTA.

5. After the high yield transcription reaction proceed with purification of synthesised RNA and/or analysis of integrity, length and yield of the transcript on agarose gel.

## Important notes

### RNase contamination

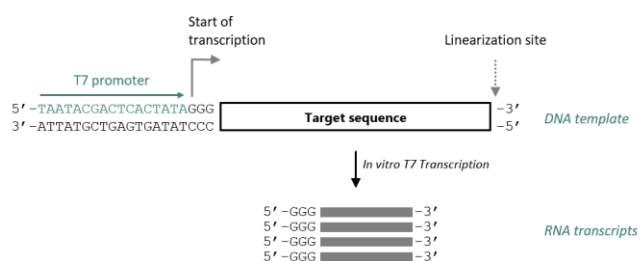
An RNase-free working environment is essential to obtain high quality RNA and the success of *in vitro* transcription. For this reason, special precautions should be taken to avoid RNase contamination, such as:

- Always wear gloves; change gloves whenever you suspect of a contamination;
- Use RNase-free tubes and pipet tips;
- Designate a special area and equipment dedicated for RNA work.

All components of NZY T7 High Yield RNA Synthesis kit were tested to ensure the absence of RNases.

### DNA Template

Any linearized plasmids or PCR products that contain a T7 promoter region may be used as template for high yield *in vitro* transcription using the NZY T7 High Yield RNA Synthesis kit. The sequence of the T7 promoter recognized by T7 RNA polymerase is indicated in Figure 1:



**Figure 1.** *In vitro* transcription from the T7 promoter.

The orientation of RNA synthesis (sense or antisense) depends on the orientation of the T7 promoter relatively to the sequence of interest. Thus, the sequence of interest must be placed downstream of the T7 promoter (for sense RNA transcripts, see Figure 1).

### Plasmid templates

The success of *in vitro* transcription using the NZY T7 High Yield RNA Synthesis kit depends on the quality and linearization of plasmid templates, since these two parameters affect the quality and quantity of synthesised RNA transcripts. Plasmid templates need to be linearized before the *in vitro* transcription reaction to allow the production of RNA transcripts with a defined length.

Since T7 RNA polymerases are highly processive enzymes, circular plasmid templates are not recommended and will generate long and heterogeneous RNA transcripts. Although the *in vitro* transcription reaction will stop when T7 RNA polymerase recognizes a loop or hairpin termination region its efficacy will depend on the type of terminator. Therefore, when using plasmid templates to generate RNAs with a defined length, it is highly recommended to digest the DNA at the 3' region where transcription is aimed to stop. Hence, it is highly recommended to linearize the plasmid template by the restriction digestion the DNA at a position located downstream of the insert to be transcribed (Figure 1).

Preferentially, the DNA template should be double-digested upstream and downstream the region that needs to be transcribed. The use of restriction enzymes which generate blunt ends or 5' overhangs are preferred related to restriction enzymes that produce 3' overhangs. If the last enzymes must be used, the linearized plasmid ends can be made blunted using Klenow Fragment of DNA polymerase I (MB009) prior to transcription.

### PCR templates

DNA templates generated by PCR and containing a T7 promoter can be effectively transcribed using the NZY T7 RNA Synthesis kit. PCR products should be examined on an agarose gel (to estimate concentration and to confirm the expected size) before their incorporation in the *in vitro* transcription reaction. We recommend purifying PCR products to obtain better transcription yields.

### Amounts of DNA template

We recommend using 0.5-1 µg of any DNA template in a 20 µL *in vitro* transcription reaction.

## Quality control assays

### Purity

T7 HY Enzyme Mix components are >95% pure as judged by SDS polyacrylamide gel electrophoresis followed by Coomassie blue staining.

### Nucleases assay

All components of the kit are tested for DNases and RNases activities, using 0.2-0.3 µg of pNZY28 plasmid DNA and 1 µg of RNA, respectively. Following incubation at 37 °C for 14-16 hours for DNases assay and 1 hour for RNases assay, nucleic acid integrity is evaluated by agarose gel electrophoresis.

### Functional assay

All components of the kit are functionally tested in an *in vitro* transcription reaction. 150 µg of intact RNA transcript must be produced from 1 µg of linearized DNA template.

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### Certificate of Analysis

Test	Result
Enzyme purity	Pass
Nucleases assay	Pass
Functional assay	Pass

Approved by:

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